

CRYOLOCK



WWW.CRYOLOCK.INFO

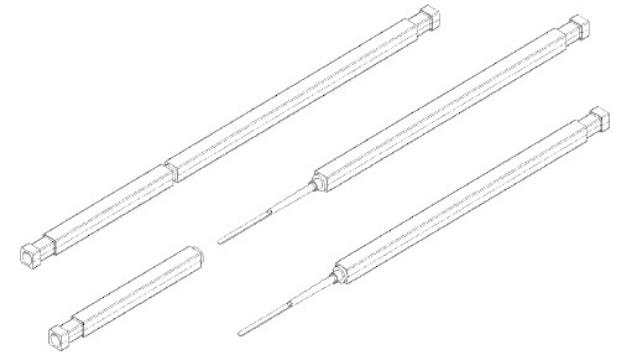
SALES@BIODISENO.COM

For more information go to:

www.Cryolock.info

Certificates of Analysis
Available upon request

Distributed by:



NEW DESIGN

Thinner for more space
available on dewars.



Available on 5 different
colors:

Orange, Blue, Green,
Yellow, and Clear

CRYOLOCK™



System

For Oocyte and/or
Embryo Vitrification



0086



EC REP

Atlantico Systems Ltd
34 Oldfield, Kingston
Galway, Ireland
+35391443609

REF

CL-R-CT

STERILE R

SAL 10⁻⁶



Biotech Inc.

5010 Ivy Nole, Cumming, GA 30040

1-800-313-7793

DO NOT
RE-USE



INSTRUCTIONS FOR USE

Cryolock™ is an assisted reproduction labware intended to be used for holding, cryopreservation and storage of oocytes or embryos under liquid nitrogen (LN₂) temperatures.

Warnings

All procedures must be performed under aseptic laboratory conditions.

To avoid injuries with LN₂, wear protective gloves and glasses.

Do not use Cryolock™ if: (a) vial or package is damaged, (b) gamma indicator is yellow or missing, or (c) Expiration Date has expired.

Before loading oocytes or embryos, verify integrity of Cryolock™ Tip.

For better survival rates, use MII Oocytes or Good Quality Embryos for vitrification.

Do not touch the tip of Cryolock™ at any time; even avoid contact with any surface.

For infectious patients take additional precautions and follow laboratory procedures for those special cases.

When plunging Cryolock™ into LN₂ always use a separate fresh aliquot LN₂ per patient.

Do not Re-sterilized or Re-use Cryolock. Device properties may change decreasing Cryolock performance. Possible contamination, low survival rates, lysis and/or Oocyte/Embryo degeneration may occur.

Precautions

The correct use of the device is responsibility of the user. For exclusive use of embryologists, biologists or laboratory technicians duly trained on cryopreservation techniques and vitrification protocols.

For vitrification and warming purposes, have all necessary materials, tools and equipment ready and handy before starting procedures.

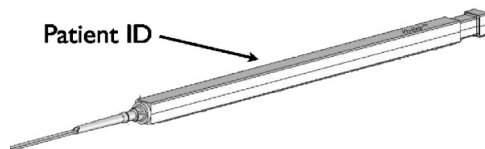
For Laboratory use only. Not for diagnostic use.

Storage: Store at room temperature

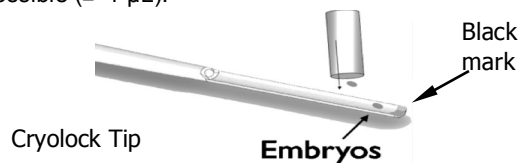
This product has not been evaluated by The FDA.
LL-5001 Rev. D – 2/23/2012 DOC#: 12-01

LOADING AND CLOSING

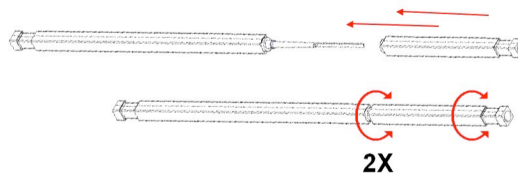
1. Use a liquid nitrogen-resistant label to identify oocytes and embryos of the patients, using the label on the same surface where Cryolock™ is engraved.



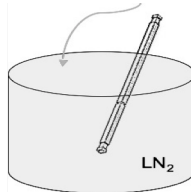
2. Prepare the sample for vitrification according to laboratory vitrification protocol.
3. Using a micropipette, carefully load a maximum of 2 specimens on the concave surface of the tip (same side of Cryolock™ logo) and about 3mm (1/8") from the edge of tip (use black mark as a reference) removing any excess of cryo-protectant solution leaving as minimum volume of vitrification media as possible ($\leq 1 \mu\text{L}$).



4. Immediately immerse Tip and Cap under LN₂. Allow equilibration until stop bubbling. Carefully insert the tip into the cap twisting tightly enough until secure.



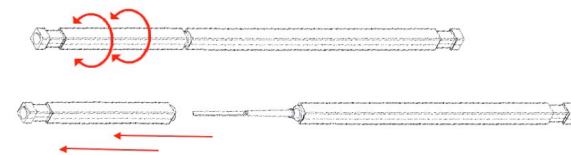
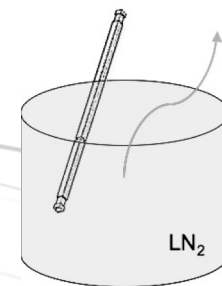
5. Storage specimens on dewars following the laboratory vitrification protocol. Always store the Cryolock™ with the cap facing down.



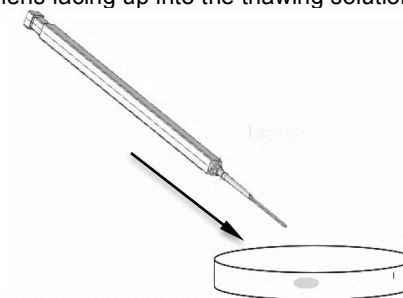
Note: Perform steps 3 to 5 in less than 1 minute. After vitrification, Cryolock™ must be kept under liquid nitrogen at all times.

WARMING

1. Prepare the thawing solutions according to laboratory vitrification protocol.
2. Identify the sample to be thawed.
3. Place the warming solution under microscopic view.
4. Using forceps hold the upper extreme of the Cryolock™ body and quickly but gently remove the cap under LN₂ twisting the parts until release.



5. Immediately plunge the tip of Cryolock™ with specimens facing up into the thawing solution at 37°C.



6. Under microscopic observation, gently move the Cryolock™ until embryos are released from the tip.
7. Continue the warming according to laboratory vitrification / warming protocol.
8. Discard Cryolock after completion of procedure.

Note: Transition between steps 4 to 5 should be no longer than 5 seconds.